See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/324522866

Development of a body condition score for the mountain chicken frog (Leptodactylus fallax)

Article *in* Zoo Biology · April 2018

CITATIONS 2	5	READS 156	
10 autho	ors, including:		
	Stephanie Jayson RSPCA Science and Policy Group 13 PUBLICATIONS 24 CITATIONS SEE PROFILE		Luke Harding Zoological Society of London 21 PUBLICATIONS 32 CITATIONS SEE PROFILE
	Christopher Michaels Zoological Society of London 55 PUBLICATIONS 242 CITATIONS SEE PROFILE		Benjamin Tapley ZSL 79 PUBLICATIONS 404 CITATIONS SEE PROFILE

Some of the authors of this publication are also working on these related projects:

EVALUATING POPULATIONS OF THE ACHOQUE (AMBYSTOMATIDAE: AMBYSTOMA DUMERILII) IN LAKE PÁTZCUARO FOR THE RECOVERY OF LOCAL MANAGEMENT AND FISHERIES View project

Mountain Chicken Recovery Programme View project

RESEARCH ARTICLE

WILEY ZOOBIOLOGY

Development of a body condition score for the mountain chicken frog (*Leptodactylus fallax*)

Stephanie Jayson^{1,2} | Luke Harding³ | Christopher J. Michaels¹ | Benjamin Tapley¹ | Joanna Hedley² | Matthias Goetz⁴ | Alberto Barbon⁴ | Gerardo Garcia⁵ | Javier Lopez⁵ | Edmund Flach¹

¹Zoological Society of London, London Zoo, Regent's Park, London

² The Royal Veterinary College, London

³ Paignton Zoo Environmental Park, Paignton, Devon

⁴ Durrell Wildlife Conservation Trust, Jersey Zoo, Trinity, Jersey

⁵ Chester Zoo, Upton-by-Chester, Upton, Chester

Correspondence

Stephanie Jayson, Veterinary Department, Zoological Society of London, London Zoo, Regent's Park, London NW1 4RY, UK. Email: stephanie.jayson@zsl.org

Funding information Royal Veterinary College The Critically Endangered mountain chicken frog (Leptodactylus fallax) has undergone drastic population decline due to habitat loss, hunting, invasive species, and chytridiomycosis. In response, several partner institutions initiated a conservation breeding program. It is important to maintain the captive population in good health. Therefore the program partners have recommended establishment of protocols for health examination of the species, including body condition assessment. Visual body condition scoring is a useful means to assess body condition in zoo animals for which regular bodyweight measurements are impractical or associated with capture-related stress. In this study, the authors developed a visual body condition score for the mountain chicken frog based on an ordinal categorical scale from 1 to 5 (1 =lowest body condition, 5 = highest body condition) using anatomical features that vary with total body energy reserves. Veterinary staff, animal managers, keepers, researchers, and students subsequently used the body condition score to assign scores to 98 mountain chicken frogs (41 male, 57 female) aged between 8 months and 12 years housed in five zoos in the UK and Jersey between February and March 2016. Body condition scores showed moderate (rho = 0.54; males) to strong (rho = 0.6; females) correlation with the scaled mass index, an objective measure of total energy reserves. The majority of pairwise comparisons between scores showed slight to substantial intra-observer agreement (93.8%) and slight to almost perfect inter-observer agreement (97.2%). Cases of poor agreement were likely due to limited observer experience working with the species.

KEYWORDS

amphibian, conservation, health assessment, zoo

1 | INTRODUCTION

The mountain chicken frog (*Leptodactylus fallax*) is a large Critically Endangered anuran native to the Caribbean islands of Dominica and Montserrat (IUCN SSC Amphibian Specialist Group, 2017). The species recently underwent one of the fastest population declines recorded, with 85% loss of the population in less than 18 months on Dominica and almost complete extirpation on Montserrat due to the fungal disease chytridiomycosis (Hudson et al., 2016). Consequently, several institutions established a captive population for conservation breeding (Adams et al., 2014; IUCN SSC Amphibian Specialist Group, 2017). The immediate future of the species on Montserrat is uncertain with the most realistic chance of success being through captive breeding and release (Adams et al., 2014).

The partners of the conservation breeding program are committed to ensuring the highest health standards are maintained. Therefore, they have recommended development of common protocols to allow comparable health assessments across the captive population, including methods for assessment of body condition (Adams et al., 2014). For the purposes of this study, body condition is defined as an individual's total body energy reserves, the measurable component of which is fat mass plus lean body mass (Clancey & Byers, 2014). As poor body condition is often reported with many diseases of amphibians, development of a method to assess body condition for use by animal husbandry and veterinary staff could improve health monitoring of species in this taxonomic group (Chai, 2015; Densmore & Green, 2007). Veterinarians commonly observe thin to emaciated body condition within the captive population of mountain chicken frogs at post mortem examination, often in specimens affected by intestinal adenocarcinoma which is frequently diagnosed in this species (Barbon, Flach, & Lopez, pers. obs.; Jaffe et al., 2015). With experience and training, regular body condition assessment may provide an earlier warning of certain diseases in the captive population (Schiffmann, Clauss, Hoby, & Hatt, 2017).

There are several possible methods to estimate body condition. The gold standard direct measure is chemical analysis of carcass tissues for lipid content (Mawby et al., 2004). However, stored whole carcasses may not be readily available and euthanasia of animals for this purpose is not usually justifiable in a zoo setting, particularly for threatened species such as the mountain chicken frog. Indirect methods include dual-energy X-ray absorptiometry (DEXA), deuterium oxide (D₂O) dilution, quantitative magnetic resonance (QMR), body condition indices based on morphometric measurements (such as the scaled mass index), and body condition scores which are based on visual assessment, with or without manual palpation, of key anatomical features (Clancey & Byers, 2014). Apart from visual body condition scores, all of these techniques require some form of manual or chemical restraint. Therefore, body condition scores based on visual assessment may be considered a more appropriate method for regular measurement of body condition of captive and free-ranging wild animals for which repeated manual or chemical restraint is impractical or associated with capture-related stress (Clements & Sanchez, 2015; Schiffmann et al., 2017; Thomson, Burkholder, Heithaus, & Dill, 2009).

Body condition scores should be: 1) Well described; 2) Relevant to the species to which they are applied; 3) Repeatable (low intraobserver variability); 4) Reproducible (low inter-observer variability); and 5) Correlate well with objective measures of body fat content (Laflamme, 1997; Summers, Clingerman & Yang, 2012). It has been recommended that body condition scores are tested for the latter three points (Schiffmann, Clauss, Hoby, & Hatt, 2017). Although body condition scores have been described in a number of captive and free ranging wild mammals, birds and reptiles, few scoring systems have been assessed for repeatability, reproducibility, and correlation with an objective measure of body fat content and to the authors' knowledge there are no published body condition scores for an amphibian species (Burton, Newnham, Bailey, & Alexander, 2014; Lamberski, 2013; Schiffmann et al., 2017; Thomson, Burkholder, Heithaus, & Dill, 2009). The aim of this study is to develop a visual body condition score (BCS) for the mountain chicken frog that could be used by animal husbandry and veterinary staff to monitor body condition of individuals of this species in captivity.

2 | MATERIALS AND METHODS

2.1 | Development of the body condition score (BCS)

The authors requested photographs of captive mountain chicken frogs at rest in their enclosures from institutions housing this species and subsequently excluded images of restrained mountain chicken frogs due to distortion of anatomical landmarks when held. The primary author, experienced mountain chicken frog keepers, and veterinary staff at the Zoological Society of London reviewed the photographic database, alongside personal observations of the species, and selected potential anatomical features for the BCS that: 1) Could be assessed visually with the animal in a standard resting position without requiring handling; 2) Were expected to vary with body energy reserves; and 3) Would not be directly affected by other variables (e.g. gender, sex-related seasonal changes, age, and posture). They initially selected the following anatomical features: a) the soft tissue covering the sacrum, suprascapulae, ilia, and urostyle; b) the crus at its widest part (compared to the maximum width of the palpebral opening); c) the dorsolateral dermal ridge; and d) the soft tissue in the gular region (Figure 1). A BCS was developed with drawings and written descriptions of the selected anatomical landmarks at different grades of condition using an ordinal categorical scale ranging from 1 (lowest condition) to 5 (highest condition). A comment on the overall shape of the body (for example, angular or rounded) was also included in the written descriptions.

2.2 | Assessment of the BCS

The authors performed a pilot study to assess for intra- and interobserver variability when observers used the BCS to score photographs of mountain chicken frogs. The primary author selected 19 photographs of captive mountain chicken frogs which included frogs representing the full range of body condition scores from 1 to 5, in a natural resting position, with the selected anatomical landmarks clearly visible. Eight observers (four veterinary surgeons, three keepers, and one veterinary nurse) were each provided with a copy of Figure 1, as well as the BCS, and the nineteen photographs which were randomly ordered. The observers assigned a score from 1 to 5 to each photograph based on the BCS. Four observers repeated the process 1 week later. Observers were asked to give written feedback on difficulties encountered when using the BCS. Observers were blinded to their previous scores and to those assigned by other observers.

Intra- and inter-observer agreement between scores was assessed using weighted κ statistics and expressed using cut-off values between 0 and 1 whereby $\kappa < 0.00$ represents poor agreement, $0.00 \le \kappa \le 0.20$ slight agreement, $0.20 < \kappa \le 0.40$ fair agreement, $0.40 < \kappa \le 0.60$ moderate agreement, $0.60 < \kappa \le 0.80$ substantial agreement and $0.80 < \kappa \le 1.00$ almost perfect agreement (Cohen, 1968; Landis & Koch, 1977). Tables

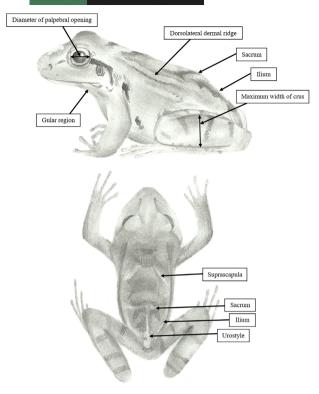


FIGURE 1 Anatomical features initially selected for use in development of the mountain chicken frog (*Leptodactylus fallax*) body condition score (BCS). The dorsolateral dermal ridge was not used in the final BCS (Figure 2) following observer feedback that it appeared to vary inconsistently with body condition

consisting of two columns containing the scores for each combination of two observers were inputted into an online calculator using the Cohen's Kappa for two raters setting and the weighted κ value reported (StatsToDo, 2014).

Observers noted in written feedback that the dorsolateral dermal ridge appeared to vary inconsistently with body condition therefore the authors removed the dorsolateral dermal ridge as an anatomical feature of the BCS, resulting in the final BCS shown in Figure 2. The authors subsequently assessed the BCS for intra- and inter-observer variability, as well as correlation with an objective measure of body condition (scaled mass index), using live mountain chicken frogs. Four to six observers at each of five zoos (A-E) used the BCS to assign a body condition score to the live mountain chicken frogs held at each zoo. The study population comprised the total population of mountain chicken frogs in captivity in the UK and Jersey, excluding juveniles under 6 months of age (nine individuals) as they were not due to be handled for bodyweight and snout-vent length measurements due to their young age. This resulted in a total study population of 98 frogs (41 male, 57 female) aged between 8 months and 12 years, including 12 frogs aged 3 years at Zoo A, 37 frogs aged 8 months-11 years at Zoo B, 14 frogs aged 4–12 years at Zoo C, eight frogs aged 2 years at Zoo D and 27 frogs aged 3 years at Zoo E. Observers scored the frogs just prior to the species' breeding season (between late February and late March 2016). Observer 5 at zoo C, and 3, 4, and 5 at zoos D and E consisted of three work experience students, a probationary keeper, a

trainee keeper, a seasonal keeper, and a conservation researcher. They were considered less experienced working with mountain chicken frogs than other observers which included full time veterinarians. veterinary nurses, keepers, and animal management staff. Three of the authors involved in designing the BCS participated as observers (Observer 1 at Zoos A-E and Observers 2 and three at Zoo A): as such these observers were more familiar with the anatomical features of the BCS. Other observers were sent a copy of Figure 1 (with the dorsolateral dermal ridge removed) and the BCS (Figure 2) 1 week prior to scoring; however, observers were not required to view the BCS prior to use. No training in use of the BCS or opportunities to practice its use with captive mountain chicken frogs were provided. Each observer viewed each mountain chicken frog at rest in their enclosure prior to handling (Zoos A, C, D, and E) or at rest in an open transparent plastic box immediately after being moved from their enclosure (Zoo B) for a period of up to five minutes and assigned a score based on the BCS. At Zoos B, C, D, and E this was repeated 24 or 48 hr later. Individual identification of the mountain chicken frogs was confirmed by scanning the electronic microchip of each individual immediately following scoring and was not known to observers at the time of scoring.

Following one scoring event for each frog, the snout-vent length (mm to nearest 1 mm) was measured by the primary author from the most rostral edge of the snout to the most proximal edge of the vent using electronic calipers (LCD Digital Electronic Caliper Vernier Gauge Micrometer Tool, LUPO, UK) with the frog in hand, held around the waist. Bodyweight (grams to nearest 1 g) was measured using electronic scales (Tare and Fine Digital Kitchen Scale, Tanita, NL) immediately following body condition scoring. The authors calculated the scaled mass index for each mountain chicken frog using the formula described by Peig and Green (2009): scaled mass index $\widehat{M}_i = M_i \left[\frac{L_i}{L_i}\right]^{b_{SMA}}$ where M_i and L_i are the body mass and the snout-vent length of individual *i* respectively; b_{SMA} is the scaling exponent estimated by the standardised major axis (SMA) regression of M on L; L_0 is the mean value of snout-vent length L for the study population; and \widehat{M}_i is the predicted body mass for individual *i* when the snout-vent length is standardized to L_0 .

The authors used Spearman rank correlation in Excel to assess for correlation between the mean body condition score assigned to an individual mountain chicken frog by all observers and its scaled mass index, and weighted κ statistic to test for agreement between body condition scores assigned by independent observers and by the same observer (Cohen, 1968; Millar, 2001; StatsToDo, 2014).

2.3 | Welfare considerations

Mountain chicken frogs are routinely weighed as part of the ongoing health monitoring of this species in captivity. To reduce unnecessary stress associated with handling, the authors chose the timing of the snout-vent length and bodyweight measurements to coincide with routine bodyweight checks of these individuals. Experienced animal husbandry and veterinary staff handled the subjects, with at least two people (one handler, one data recorder) present to keep the time

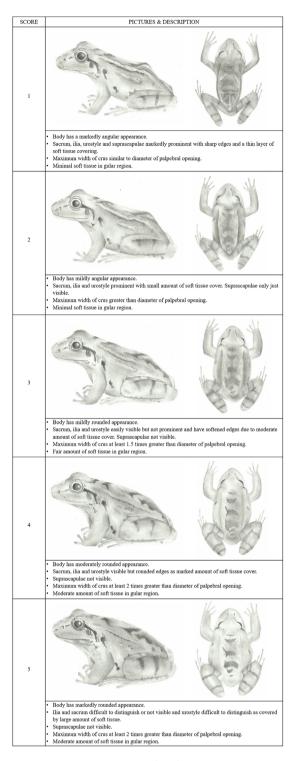


FIGURE 2 Body condition score (BCS) for the mountain chicken frog (*Leptodactylus fallax*)

in-hand to a minimum. Handlers wore moistened powder-free nitrile gloves to minimize the risk of skin damage during handling (Wright, 2001). Research proposal forms were completed, reviewed, and authorized by each institution as required by the institution's individual research department. As the procedures involved were part of routine husbandry and management procedures for the species, the institutions did not require formal ethical review.

3 | RESULTS

3.1 | Pilot study using the initial BCS and photographs

All pairwise comparisons of scores recorded by the same observer for the same photograph of a mountain chicken frog at different times (intra-observer agreement) were substantial to almost perfect (0.60 < $\kappa \le 1.00$). The majority (92.9%) of pairwise comparisons of scores recorded by different observers of the same photograph of a mountain chicken frog at the same time (inter-observer agreement) were fair to substantial (0.20 < $\kappa \le 0.80$) and 7.1% showed slight agreement (0.00 ≤ $\kappa \le 0.20$).

-Wiley-<mark>ZOO</mark>BIOLOGY

3.2 | Live animal study using the final BCS

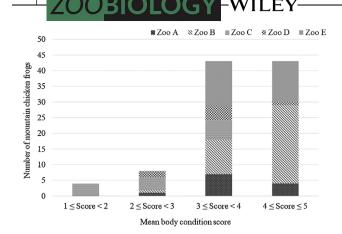
Four of the scores assigned to mountain chicken frogs at Zoo D were excluded from analysis (one by Observer 3 and three by Observer 4) as half scores, either 2.5 or 3.5, had been assigned. One frog at Zoo C was not handled to obtain snout-vent length and bodyweight measurements as it had a pre-existing leg fracture.

The distribution of mean body condition scores assigned using the BCS to the captive population of mountain chicken frogs aged between 8 months and 12 years in the UK and Jersey just prior to the breeding season in 2016 is shown in Figure 3. The majority (88%) of captive mountain chicken frogs had a mean body condition score between 3 and 5, with only 12% of individuals having a mean body condition score <3.

κ values for each pairwise comparison of the scores assigned to live mountain chicken frogs by the same observer (intra-observer agreement) and different observers (inter-observer agreement) are shown in Tables 1 and Table 2,. Intra-observer agreement was slight to substantial (0.00 ≤ κ ≤ 0.80) for 93.8% of pairwise comparisons, while 6.25% of pairwise comparisons showed poor agreement (κ < 0.00). Inter-observer agreement was slight to almost perfect (0.00 ≤ κ ≤ 1.00) for 97.2% of pairwise comparisons, with 2.78% showing poor agreement (κ < 0.00). Most scores assigned by two different observers to the same frog on the same day were within 1 score of each other, with only 4.22% of scores assigned being >1 score apart (Figure 4).

When the authors removed the less experienced observers' scores from the dataset (Observer 5 at zoo C, and 3, 4, and 5 at zoos D and E), intra- and inter-observer agreement improved such that 100% of κ values for pairwise comparisons of scores recorded by the same observer were slight to substantial ($0.20 < \kappa \le 0.8$) and 100% of pairwise comparisons of scores recorded by different observers were slight to almost perfect ($0.20 < \kappa \le 1.00$). The proportion of scores within one score of each other assigned by two different observers to the same frog on the same day also improved, with only 0.70% of scores assigned being >1 score apart.

The effect of age of the frogs on agreement between scores was evaluated at Zoo B as this was the only zoo which housed both juvenile (individuals aged less than 3 years) and adult (individuals aged greater than 3 years) frogs. For the 17 juveniles, there was fair to moderate



200

FIGURE 3 Distribution of mean body condition scores assigned to the captive mountain chicken frog population aged between 8 months and 12 years in the UK and Jersey in February and March 2016 using the body condition score (BCS) in Figure 2

 $(0.20 < \kappa \le 0.60)$ inter-observer agreement for 50% of pairwise comparisons and no to slight $(0.00 \ge \kappa \le 0.20)$ inter-observer agreement for 50% of pairwise comparisons. Inter-observer agreement was far higher for the 20 adults, with moderate to substantial agreement $(0.40 < \kappa \le 0.80)$ for 91.7% of pairwise comparisons and fair agreement $(0.20 < \kappa \le 0.40)$ for 8.33% of pairwise comparisons. Intra-observer agreement showed a similar pattern, with no to moderate agreement $(0.00 \ge \kappa \le 0.60)$ for juveniles, versus moderate to almost perfect agreement $(0.40 < \kappa \le 1.00)$ for adults. For both juveniles and adults, all scores assigned by two different observers to the same frog on the same day were within one score of each other.

In the whole study population, mean body condition score showed moderate (rho = 0.54; males) to strong (rho = 0.6; females) positive correlation with scaled mass index. Mean body condition score was more strongly correlated with scaled mass index in juveniles than

TABLE 1 κ values to show intra-observer agreement when the body condition score (BCS) in Figure 2 was used to assign body condition scores to the captive mountain chicken frog population aged between 8 months and 12 years housed at four zoos in the UK and Jersey in February and March 2016

	Observer							
Zoo	1	2	3	4	5			
В	0.70	0.48	0.32	0.57				
С	0.77	0.50	0.52					
D	0.42	0.20	-0.25	0.09				
E	0.51	0.29	0.04	0.16	0.39			

 $\kappa < 0.00$ represents poor agreement, $0.00 \le \kappa \le 0.20$ slight agreement, $0.20 < \kappa \le 0.40$ fair agreement, $0.40 < \kappa \le 0.60$ moderate agreement, $0.60 < \kappa \le 0.80$ substantial agreement, and $0.80 < \kappa \le 1.00$ almost perfect agreement (Landis & Koch, 1977). Observer 1 was the same person at all zoos. Observers 2–5 were different people at each zoo. Observers 3 and 4 at zoo D and 3, 4, and 5 at zoo E were less experienced working with mountain chicken frogs and included two work experience students, a probationary keeper, a trainee keeper and a conservation researcher. All other observers were full time veterinarians, veterinary nurses, keepers, and animal management staff.

adults in both sexes (rho in males = 0.67 (strong) in juveniles, 0.44 (moderate) in adults; rho in females = 0.91 (very strong) in juveniles, 0.55 (moderate) in adults).

4 | DISCUSSION

In this study a body condition score (BCS) was developed for the mountain chicken frog (*Leptodactylus fallax*) and assessed for intra- and inter-observer variability, as well as correlation with an objective measure of body condition (scaled mass index), in February and March 2016 just prior to the beginning of the breeding season. To the author's knowledge this is the first BCS to be developed and assessed for any amphibian species.

Historically, herpetologists have determined body condition in amphibians using body condition indices based on morphometric measurements, typically snout-vent length, and bodyweight (Băncilă, Hartel, Plăiaşu, Smets, & Cogălniceanu, 2010; Bell, Carver, Mitchell, & Pledger, 2004; Denoël, Hervant, Schabetsberger, & Joly, 2002; Gendron et al., 2003; Leary, Jessop, Garcia, & Knappa, 2004; MacCraken & Stebbings, 2012; Pope & Matthews, 2002; Yahnke, Grue, Hayes, & Troiano, 2012). Fulton's index (based on the formula: $k = \frac{M}{L^3}$ where k = Fulton's factor, M= body mass, and L = length) has been used in leopard frogs (Rana pipiens) to assess the effect of an agricultural pesticide on lungworm infection (Gendron et al., 2003; Peig & Green, 2010). Relative condition, relative mass and the residual index all use ordinary least squares regression of the linearized power equation relating mass and length and one or more of these methods have been used in the mountain yellow-legged frog (Rana muscosa), paedomorphic, and metamorphic Alpine newts (Triturus alpestris), and the yellow-bellied toad (Bombina variegata) (Băncilă et al., 2010; Denoël et al., 2002; Pope & Matthews, 2002). More recently, Peig and Green (2010) favored the scaled mass index over other indices as it is not affected by the change in relationship between mass and snoutvent length as growth occurs. In one study, MacCracken and Stebbings (2012) validated the scaled mass index in larval and juvenile bullfrogs (Lithobates catesbeianus) and rough-skinned newts (Taricha granulosa) by correlation with scaled fat mass and scaled lean mass values obtained via carcass analysis. A key advantage of using the BCS of this study over body condition indices based on morphometric measurements is that manual restraint is not required. There have been anecdotal reports of reduced food intake and reduced weight gain following manual restraint for husbandry and veterinary procedures in captive mountain chicken frogs and the frequency of handling has been suggested as a potential factor that may be affecting breeding success within the captive population (Harding, Michaels, & Tapley pers. obs.). Handling has been associated with increased corticosterone levels in a number of anurans and handling-related stress for 5, 15, or 30 min was associated with decreased testosterone excretion in non-breeding male cane toads (Rhinella marina) (Narayan, Hero, & Cockrem, 2012; Narayan, Molinia, Christi, Morley, & Cockrem, 2010). Using the BCS developed in this study to monitor body condition of the captive mountain chicken frog population during the breeding season

TABLE 2 κ values to show inter-observer agreement when the body condition score (BCS) in Figure 2 was used to assign body condition scores to the captive mountain chicken frog population aged between 8 months and 12 years housed at five zoos in the UK and Jersey in February and March 2016

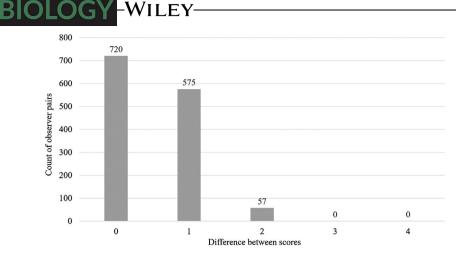
		Observer				
Zoo	Observer	2	3	4	5	6
А	1	0.17	0.39	0.80		
	2		0.17	0.17		
	3			0.39		
В	1	0.41, 0.69	0.36, 0.57	0.36, 0.44		
	2		0.50, 0.53	0.31, 0.34		
	3			0.44, 0.34		
С	1	0.93, 0.76	0.35, 0.5	0.54	0.34	
	2		0.31, 0.57	0.61	0.65	
	3			0.45	0.49	
	4				0.69	
D	1	0.20, 0.56	0.38, 0.40	0.59, 0.40	0.55	0.38
	2		0.20, 0.25	0.53, 0.75	-0.11	0.20
	3			0.70, 0.50	0.62	0.47
	4				0.17	0.30
	5					1.00
Е	1	0.44, 0.63	0.17, 0.15	0.26, 0.01	0.35, 0.27	
	2		0.19, 0.10	0.14, -0.01	0.53, 0.13	
	3			0.06, 0.06	0.09, 0.03	
	4				0.09, 0.03	

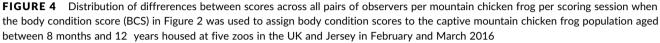
 $\kappa < 0.00$ represents poor agreement, $0.00 \le \kappa \le 0.20$ slight agreement, $0.20 < \kappa \le 0.40$ fair agreement, $0.40 < \kappa \le 0.60$ moderate agreement, $0.60 < \kappa \le 0.80$ substantial agreement and $0.80 < \kappa \le 1.00$ almost perfect agreement (Landis & Koch, 1977). Where two scores were recorded by an observer 24–48 hr apart, κ values for pairwise comparisons between observers' scores on each day are shown as x, y where x is at 0 hr and y at 24–48 hr. Observer 1 was the same person at all zoos. Observers 2–5 were different people at each zoo. Observer 5 at zoo C, and 3, 4, and 5 at zoos D and E were less experienced working with mountain chicken frogs and included three work experience students, a probationary keeper, a trainee keeper, a seasonal keeper, and a conservation researcher. All other observers were full time veterinarians, veterinary nurses, keepers, and animal management staff.

instead of manual restraint for snout-vent length and weight measurements may reduce the effects of handling-related stress during this critical time.

The authors developed an overview format of visual BCS in this study, in which observers assign a score based on overall appearance, in contrast to a composite BCS in which observers score individual body regions and calculate a sum or an algorithm BCS in which observers follow a flow chart to assign a score (Schiffmann et al., 2017). The authors preferred the overview format in this case due to its practicality and simplicity (Schiffmann et al., 2017). When developing a BCS, researchers typically select anatomical features based on appraisal of photographs of the species, advice from experienced keepers, nutritionists or veterinary medical staff, and adaptation of previously published body condition scores in the same or closely related species if available (Audigé, Wilson, & Morris, 1998; Cook et al., 2001; Ezenwa, Jolles, & O'Brien, 2009; Franzmann, 1977; German et al., 2006; Morfeld, Lehnhardt, Alligood, Bolling, & Brown, 2014; Pettis et al., 2004; Reppert, Treiber, & Ward, 2011; Rudman & Keiper, 1991; Schiffmann et al., 2017; van der Jeugd & Prins, 2000; Wemmer et al., 2006). As a BCS had not been described in similar species, the

authors selected anatomical landmarks based on the experience of keepers and veterinary staff and appraisal of photographs of the species. Intra-coelomic fat bodies are well described as a fat storage site in anurans (Pond, 1978); however, there is little published literature regarding the relative importance of externally visible fat storage sites in anurans and how they vary at different grades of condition. Amphibians lack subcutaneous fat; however fat storage has been demonstrated in the somatic musculature (Pond, 1978). The variation in anatomical sites selected for the BCS in this study with body condition likely reflect changes in energy reserves in skeletal musculature (Pond, 1978). The authors selected anatomical features that were considered to not be directly affected by other variables (e.g. gender, sex-related seasonal changes, age, and posture). For example, male mountain chicken frogs, like males of some other species of the family Leptodactylidae, develop forelimb muscular hypertrophy in the breeding season (Tapley, Acosta-Galvis, & Lopez, 2011), therefore the authors did not use forelimb size as a site in the BCS. The BCS may therefore be advantageous over body condition indices based on mass and length as these two measures may be influenced by factors other than energy reserves such as age, sex, developmental stage, hydration





status, gut fill, structural deformities, and reproductive status (MacCracken & Stebbings, 2012). However, scores assigned using the BCS would likely still be affected by major structural deformities of the key anatomical landmarks, for example, caused by trauma or metabolic bone disease, although the latter is no longer a common problem in the captive population as mountain chicken frogs are routinely fed a variety of invertebrates supplemented with a high-calcium multivitamin and mineral supplement containing vitamin D_3 and provided with appropriate levels of UV-B radiation (Tapley et al., 2014).

Evaluation of body condition scores for intra- and interobserver variability is important given the subjective nature of these systems, resulting in variability in scores assigned by independent observers (Clancey & Byers, 2014). The BCS of this study was assessed for intra- and inter-observer agreement in a similar way to BCS assessment in other species, with multiple observers assigning condition scores at least twice and a length of time apart which would be unlikely to result in a change in condition (Burton et al., 2014; Clingerman & Summers, 2005; Kristensen et al., 2006; Morfeld et al., 2014). In a previous study evaluating the effect of training on intra- and inter-observer agreement between body condition scores assigned to Holstein dairy cattle by practicing dairy veterinarians, the authors showed that veterinarians who had received a 2 hr theoretical lecture regarding use of a BCS prior to its use had slight to substantial inter-observer agreement ($\kappa = 0.17 - 0.78$) (Kristensen et al., 2006). Agreement improved to moderate to substantial after a further 2.5 hr practical training session ($\kappa = 0.41 - 0.82$) and intraobserver agreement between the first and second scoring sessions was fair to substantial ($\kappa = 0.22 - 0.75$). Far higher agreement was achieved when highly trained instructors who had worked closely together in a formal network over at least 3 years used the same BCS ($\kappa \ge 0.86$) (Kristensen et al., 2006). Observer experience also appears important in other species, for example, Pettis et al. (2004) found almost perfect inter-observer agreement (κ = 0.86–0.87) when a 3-point visual BCS was applied

by three experienced right whale biologists, one of whom developed the BCS, to photographs of North Atlantic right whales, and Morfeld et al. (2014) demonstrated stronger interobserver agreement between two observers who developed a five-point visual BCS for female African elephants than between these observers and an observer with no prior experience of using a BCS (κ = 0.89 compared to 0.62–0.67). Given that no specific theoretical or practical training in use of the BCS was provided in this study but observers had the opportunity to view the BCS in advance, the authors predicted that intra- and inter-observer agreement for veterinarians, veterinary nurses, animal managers, and keepers working with the species would be slight to substantial, as Kristensen et al. (2006) observed with practicing dairy veterinarians scoring Holstein dairy cattle after a 2 hr theoretical lecture. As some of the observers in this study had no or minimal experience working with mountain chicken frogs, the authors expected intra- and inter-observer agreement to be lower than that observed by Kristensen et al. (2006) for practicing veterinarians working with dairy cattle (i.e. poor to slight). Slight to substantial agreement was achieved in 93.8% of intra-observer pairwise comparisons and slight to almost perfect agreement in 97.2% of inter-observer pairwise comparisons in this study. When the authors removed individuals with no or minimal prior experience working with mountain chicken frogs (work experience students, a conservation researcher, a probationary keeper, and a seasonal keeper) agreement increased, such that 100% of pairwise comparisons showed slight to substantial intra-observer agreement and 100% of pairwise comparisons showed slight to almost perfect inter-observer agreement. When using a BCS in other species, agreement between scores can be improved with training and experience and it has become a useful component of routine clinical examination to estimate body energy reserves in many species (Bewley & Schutz, 2008; Clancey & Byers, 2014; Clements & Sanchez, 2015; Houston & Radostits, 2000, Kristensen et al., 2006). With further training in use of the mountain chicken frog BCS, the authors expect that agreement between scores assigned at zoos with less experienced staff would improve over time.

The second stage of assessment is correlation with an objective measure of total energy reserves. The gold standard objective measure is carcass analysis (Gerhart, White, Cameron, & Russell, 1996). However, the authors did not consider euthanasia for carcass analysis following condition scoring appropriate for the mountain chicken frog given the threatened status of the species and the importance of animals involved in this study for maintaining the captive population (Adams et al., 2014). Researchers may use other indirect objective measures such as dual-energy X-ray absorptiometry (DEXA), deuterium oxide (D₂O), dilution, and quantitative magnetic resonance imaging (QMR) in species for which these techniques have been validated; however, none of these methods have been validated in the mountain chicken frog or any amphibian species (Laflamme, 1997; Nixon et al., 2010; Rudolph, Stahly, & Cromwell, 1988). An alternative option used in certain mammalian species is measurement of subcutaneous fat by ultrasonography; however, anurans do not typically store fat subcutaneously (Pond, 1978; Schiffmann et al., 2017). Measurement of intra-coelomic fat bodies by ultrasonography could be considered; however, they can be difficult to differentiate on coelomic ultrasonography (Pond, 1978; Schildger, 2001). Therefore, the authors considered correlation with a body condition index based on morphometric measurements which had been validated in other amphibian species to be the most appropriate method to validate the condition score in this study. The authors used the scaled mass index as it is not affected by changes in snout-vent length and bodyweight as growth occurs (MacCracken & Stebbings, 2012; Peig & Green, 2009, 2010). In future studies, the authors recommend validation of both the BCS developed in this study and the scaled mass index by analysis of carcass energy reserves following use of the BCS and measurement of scaled mass index as carcasses become available.

The effect of age on agreement between scores was evaluated at Zoo B which housed 17 juvenile and 20 adult frogs. There was no to moderate intra- and inter-observer agreement for juvenile frogs, whereas there was moderate to almost perfect intra-observer agreement and fair to substantial inter-observer agreement for adults. Clingerman and Summers (2012) also found inter-observer agreement was poorer in juveniles than adults when a BCS was used in rhesus macaques, likely due to there being less muscle mass and fat reserves in normal juveniles, and juveniles may be experiencing growth and changes in body stature. Despite the apparent poorer agreement between scores assigned to juvenile frogs at Zoo B, all of the scores assigned to juvenile frogs by different observers on the same day at Zoo B were no greater than one score apart and correlation with the scaled mass index was strong to very strong in the total study population of juvenile frogs. Further evaluation of the BCS in juvenile frogs is recommended to determine its utility in this age group given the small number of juvenile frogs within this study population.

The authors assessed the BCS in this study at one time-point of the year, just prior to the breeding season. Previous studies with

other species such as the Magellanic penguin (*Spheniscus magellanicus*) have demonstrated variation in BCS with season (Clements & Sanchez, 2015). Many anuran species, including the mountain chicken frog, expend a great deal of energy by engaging in reproductive behavior due to egg production, and parental care in females and vocalization and agonistic behavior in males (Fitzpatrick, 1976; Gibson & Buley, 2004; Mizell, 1965; Morton, 1981; Seymour, 1973; Smith, 1950). These changes will likely change the distribution of body condition scores within a population. Therefore the authors recommend that the mountain chicken frog BCS is evaluated with data collected at different times of the year, for example at the end of the breeding season once energy resources are most depleted.

-WILEY-700BIOLOG

The BCS of this study represents the range of body condition observed in the captive mountain chicken frog population from 1 (lowest condition) to 5 (highest condition). In mammals and birds, the lowest body condition score of a BCS is typically described as emaciated, the highest as obese, and the middle score considered normal or ideal (Bewley & Schutz, 2008; Clements & Sanchez, 2015; Laflamme, 1997). However, the ideal score depends on the context, for example in dairy cattle the ideal body condition score depends on the stage of lactation and the production system (Bewley & Schutz, 2008). Descriptive terms such as emaciated, normal, and obese were not assigned to the numerical scores in this study as insufficient information is known about which grade of condition constitutes normal. Obesity has been described in certain species of frog, such as the White's tree frog (Pelodryas caerulea), and is characterized by abdominal distension due to fat deposition in the coelomic fat bodies and enlargement of the supraocular skin folds in this species, which may impair vision (Wright & Whitaker, 2001). However clinical obesity, defined as fat accumulation which may impair health, has not been reported to date in captive mountain chicken frogs (Lopez, pers. obs.). In contrast, thin to emaciated body condition is commonly observed at post mortem examination (Barbon, Flach & Lopez, pers. obs). The majority (88%) of the mountain chicken frogs in the captive population in this study had a mean body condition score of 3-5 just prior to the breeding season, which likely represents normal condition, while 12% had a mean score of <3 which is likely abnormal, with two representing thin and one emaciated body condition. The authors recommend correlation between body condition scores assigned using the BCS and health parameters, as well as other parameters important to conservation breeding programs, such as reproductive output, longevity and post-release survival, to determine the ideal body condition of this species in different contexts (Bewley & Schutz, 2008; Pettis et al., 2004).

5 | CONCLUSIONS

- 1) A body condition score (BCS) has been developed for the mountain chicken frog (*Leptodactylus fallax*).
- 2) The BCS has been evaluated in February-March just prior to the beginning of the breeding season. Agreement between scores

assigned by the same and different observers is expected to be slight to substantial if observers have received no formal training with the BCS but have experience working with mountain chicken frogs. Agreement between scores may be poor if observers have received no formal training with the BCS and have no or minimal experience working with mountain chicken frogs.

3) Further assessment of the BCS is recommended for use in juveniles and in adults at other times of year, for example, at the end of the breeding season when energy supplies are likely to be most depleted.

ACKNOWLEDGMENTS

The authors would like to thank the staff at Bristol Zoo, Chester Zoo, Chessington Zoo, Jersey Zoo, and the Zoological Society of London who contributed to data collection. Thank you to Ruby Chang (Royal Veterinary College) for advice on statistical analysis and to the Royal Veterinary College for providing funding for this research.

ORCID

Stephanie Jayson D http://orcid.org/0000-0003-4714-0417 Christopher J. Michaels D http://orcid.org/0000-0002-4733-8397

REFERENCES

- Adams, S. L., Morton, M. N., Terry, A., Young, R. P., Dawson, J., Martin, L., ... Gray, G. (2014). Long-term recovery strategy for the critically endangered mountain chicken 2014–2034. Retrieved from http:// www.amphibians.org/wp-content/uploads/2015/08/Mountain-Chick en-SAP-2014-working-draft-FINAL.pdf
- Audigé, L., Wilson, P. R., & Morris, R. S. (1998). A body condition score system and its use for farmed red deer hinds. New Zealand Journal of Agricultural Research, 41, 545–553.
- Băncilă, R. I., Hartel, T., Plăiaşu, R., Smets, J., & Cogălniceanu, D. (2010). Comparing three body condition indices in amphibians: A case study of yellow-bellied toad Bombina variegata. Amphibia-Reptilia, 31, 558–562.
- Bell, B. D., Carver, S., Mitchell, N. J., & Pledger, S. (2004). The recent decline of a New Zealand endemic: How and why did populations of Archey's frog *Leiopelma archeyi* crash over 1996-2001? *Biological Conservation*, 120, 193–203.
- Bewley, J. M., & Schutz, M. M. (2008). Review: An interdisciplinary review of body condition scoring for dairy cattle. *The Professional Animal Scientist*, 24, 507–529.
- Burton, E. J., Newnham, R., Bailey, S. J., & Alexander, L. G. (2014). Evaluation of a fast, objective tool for assessing body condition of budgerigars (*Melopsittacus undulatus*). Journal of Animal Physiology and Animal Nutrition, 98, 223–227.
- Chai, N. (2015). Anurans. Fowler's zoo and wild animal medicine (8th ed., pp. 1–13). Missouri: Elsevier Saunders.
- Clancey, E., & Byers, J. A. (2014). The definition and measurement of individual condition in evolutionary studies. *Ethology*, 120, 845–854.
- Clements, J., & Sanchez, J. N. (2015). Creation and validation of a novel body condition scoring method for the magellanic penguin (Spheniscus magellanicus) in the zoo setting. Zoo Biology, 34, 538–546.
- Clingerman, K. J., & Summers, L. (2005). Development of a body condition scoring system for nonhuman primates using *Macaca mulatta* as a model. *Lab Animal*, 34, 31–36.

- Cohen, J. (1968). Weighted kappa: Nominal scale agreement with provision for scaled disagreement or partial credit. *Psychological Bulletin*, 70, 213–220.
- Cook, R. C., Cook, J. G., Murray, D. L., Zager, P., Johnson, B. K., & Gratson, L. W. (2001). Development of predictive models of nutritional condition for rocky mountain elk. *The Journal of Wildlife Management*, 65, 973–987.
- Denoël, M., Hervant, F., Schabetsberger, R., & Joly, P. (2002). Short- and long-term advantages of an alternative ontogenetic pathway. *Biological Journal of the Linnean Society*, 77, 105–112.
- Densmore, C. L., & Green, D. E. (2007). Diseases of amphibians. ILAR Journal, 48, 235–354.
- Ezenwa, V. O., Jolles, A. E., & O'Brien, M. P. (2009). A reliable body condition scoring technique for estimating condition in African buffalo. *African Journal of Ecology*, 47, 476–481.
- Fitzpatrick, L. C. (1976). Life history patterns of storage and utilization of lipids for energy in amphibians. *American Zoologist*, 16, 725–732.
- Franzmann, A. W. (1977, April). Condition assessment of Alaskan moose. Paper session presented at the North American Moose Conference and Workshop, Jasper, Alberta.
- Gendron, A. D., Marcogliese, D. J., Barbeau, S., Christin, M. S., Brousseau, P., Ruby, S., . . . Fournier, M. (2003). Exposure of leopard frogs to a pesticide mixture affects life history characteristics of the lungworm *Rhabdias ranae*. *Oecologia*, 135, 469–476.
- Gerhart, K. L., White, R. G., Cameron, R. D., & Russell, D. E. (1996). Estimating fat content of caribou from body condition scores. *The Journal of Wildlife Management*, 60, 713–718.
- German, A. J., Holden, S. L., Moxham, G. L., Holmes, K. L., Hackett, R. M., & Rawlings, J. M. (2006). A simple, reliable tool for owners to assess the body condition of their dog or cat. *Journal of Nutrition*, 136, 2031S–2033S.
- Gibson, R. G., & Buley, K. R. (2004). Maternal care and obligatory oophagy in Leptodactylus fallax: A new reproductive mode in frogs. Copeia, 2004, 128–135.
- Houston, D. M., & Radostits, O. M., (2000). The clinical examination. In O. M. Radostits, I. G. Mayhew, & D. M. Houston (Eds.), Veterinary clinical examination and diagnosis (1st ed., pp. 91–124). London: Elsevier.
- Hudson, M. A., Young, R. P., D'Urban Jackson, J., Orozco-terWengel, P., Martin, L., James, A., ... Cunningham, A. A. (2016). Dynamics and genetics of a disease-driven species decline to near extinction: Lessons for conservation. *Scientific Reports*, *6*, 30772.
- IUCN SSC Amphibian Specialist Group. (2017). *Leptodactylus fallax*. Retrieved from http://www.iucnredlist.org/details/57125/0
- Jaffe, J., Flach, E. J., Feltrer, Y., Rivers, S., Lopez, F. J., & Cunningham, A. A. (2015). Intestinal adenocarcinoma in a Montserrat mountain chicken (Leptodactylus fallax). Journal of Zoo and Aquarium Research, 3, 21–24.
- Kristensen, E., Dueholm, L., Vink, D., Andersen, J. E., Jakobsen, E. B., Illum-Nielsen, S., ... Enevoldsen, C. (2006). Within- and across-person uniformity of body condition scoring in Danish Holstein cattle. *Journal of Dairy Science*, 89, 3721–3728.
- Laflamme, D. (1997). Development and validation of a body condition score system for dogs. *Canine Practice*, 22, 10–15.
- Lamberski, N. (2013). Body condition scores for desert tortoises. Retrieved from https://www.fws.gov/nevada/desert_tortoise/docu ments/reports/2013/assess/Desert-Tortoise-BCS-2013-lamberskipo.pdf
- Landis, J. R., & Koch, G. G. (1977). The measurement of observer agreement for categorical data. *Biometrika*, 33, 159–174.
- Leary, C. J., Jessop, T. S., Garcia, A. M., & Knappa, R. (2004). Steroid hormone profiles and relative body condition of calling and satellite toads: Implications for proximate regulation of behavior in anurans. *Behavioral Ecology*, 15, 313–320.
- MacCracken, J. G., & Stebbings, J. L. (2012). Test of a body condition index with amphibians. *Journal of Herpetology*, 46, 346–350.
- Mawby, D. I., Bartges, J. W., d'Avignon, A., Laflamme, D. P., Moyers, T. D., & Cottrell, T. (2004). Comparison of various methods for estimating body fat in dogs. *Journal of the American Animal Hospital Association*, 40, 109–114.



- Millar, N. (2001). Biology statistics made simple using Excel. *School Science Review*, 83, 23–34.
- Mizell, S. (1965). Seasonal changes in energy reserves in the common frog, Rana pipiens. Journal of Cellular Physiology, 66, 251–258.
- Morfeld, K. A., Lehnhardt, J., Alligood, C., Bolling, J., & Brown, J. L. (2014). Development of a body condition scoring index for female african elephants validated by ultrasound measurements of subcutaneous fat. *PLoS ONE*, *9*, 1–9.
- Morton, M. L. (1981). Seasonal changes in total body lipid and liver weight in the yosemite toad. *Copeia*, 1981, 234–238.
- Narayan, E., Hero, J.-M., & Cockrem, J. F. (2012). Inverse urinary corticosterone and testosterone responses to different durations of restraint in the cane toad (*Rhinella marina*). *General and Comparative Endocrinology*, 179, 345–349.
- Narayan, E., Molinia, F., Christi, K., Morley, C., & Cockrem, J. (2010). Urinary corticosterone metabolite responses to capture, and annual patterns of urinary corticosterone in wild and captive endangered Fijian ground frogs (*Platymantis vitiana*). Australian Journal of Zoology, 58, 189–197.
- Nixon, J. P., Zhang, M., Wang, C., Kuskowski, M. A., Novak, C. M., Levine, J. A., ... Kotz, C. M. (2010). Evaluation of a quantitative magnetic resonance imaging system for whole body composition analysis in rodents. *Obesity*, *8*, 1652–1659.
- Peig, J., & Green, A. J. (2009). New perspectives for estimating body condition from mass/length data: The scaled mass index as an alternative method. *Oikos*, 118, 1883–1891.
- Peig, J., & Green, A. J. (2010). The paradigm of body condition: A critical reappraisal of current methods based on mass and length. *Functional Ecology*, 24, 1323–1332.
- Pettis, H. M., Rolland, R. M., Hamilton, P. K., Brault, S., Knowlton, A. R., & Kraus, S. D. (2004). Visual health assessment of North Atlantic right whales (*Eubalaena glacialis*) using photographs. *Canadian Journal of Zoology*, 82, 8–19.
- Pond, C. M. (1978). Morphological aspects and the ecological and mechanical consequences of fat deposition in wild vertebrates. *Annual Review of Ecology, Evolution and Systematics*, 9, 519–570.
- Pope, K. L., & Matthews, K. (2002). Influence of anuran prey on the condition and distribution of *Rana muscosa* in the sierra Nevada. *Herpetologica*, 58, 354–363.
- Reppert, A., Treiber, K., & Ward, A. (2011). Body condition scoring in cheetah (Acinonyx jubatus) advancements in methodology and visual tools for assessment. Paper Presented at the Ninth Conference on Zoo and Wildlife Nutrition, Kansas City, Missouri, Abstract retrieved from https://nagonline.net/2410/body-condition-scoring-cheetah-acinony x-jubatus-advancements-methodology-visual-tools-assessment/
- Rudolph, B. C., Stahly, T. S., & Cromwell, G. L. (1988). Estimation of body composition of neonatal pigs via deuterium oxide dilution: Validation of technique. *Journal of Animal Science*, 66, 53–61.
- Rudman, R., & Keiper, R. R. (1991). The body condition of feral ponies on Assateague Island. Equine Veterinary Journal, 23, 453–456.
- Schiffmann, C., Clauss, M., Hoby, S., & Hatt, J. M. (2017). Visual body condition scoring in zoo animals—composite, algorithm and overview

approaches in captive Asian and African elephants. Journal of Zoo and Aquarium Research, 5, 1–10.

Schildger, B. (2001). Ultrasonography in amphibians. Seminars in Avian and Exotic Pet Medicine, 10, 169–173.

700BI

WILEY-

- Seymour, R. S. (1973). Energy metabolism of dormant spadefoot toads (Scaphiopus). Copeia, 1973, 435–445.
- Smith, C. L. (1950). Seasonal changes in blood sugar, fat body, liver glycogen, and gonads in the common frog, *Rana temporaria*. Journal of Experimental Biology, 26, 412–429.
- StatsToDo. (2014). Kappa (Cohen and Fleiss) for ordinal data program. Retrieved from https://www.statstodo.com/CohenFleissKappa_Pgm.php
- Summers, L., Clingerman, K. J., & Yang, X. (2012). Validation of a body condition scoring system in rhesus macaques (*Macaca mulatta*): Assessment of body composition by using dual-energy X-ray absorptiometry. Journal of the American Association for Laboratory Animal Science, 51, 88–93.
- Tapley, B., Acosta-Galvis, A. R., & Lopez, J. (2011). A field method for sampling blood of male anurans with hypertrophied limbs. *Phyllomedusa*, 10, 75–77.
- Tapley, B., Rendle, M., Baines, F. M., Goetz, M., Bradfield, K. S., Rood, D., ... Routh, A. (2014). Meeting ultraviolet B radiation requirements of amphibians in captivity: A case study with mountain chicken frogs (*Leptodactylus fallax*) and general recommendations for pre-release health screening. *Zoo Biology*, 34, 46–52.
- Thomson, J. A., Burkholder, D., Heithaus, M. R., & Dill, L. M. (2009). Validation of a rapid visual-assessment technique for categorizing the body condition of green turtles (*Chelonia mydas*) in the field. *Copeia*, 2, 251–255.
- van der Jeugd, H. P., & Prins, H. H. T. (2000). Movements and group structure of giraffe (*Giraffa camelopardalis*) in Lake Manyara National Park, Tanzania. Journal of Zoology, 251, 15–21.
- Wemmer, C., Krishnamurthy, V., Shrestha, S., Hayek, L.-A., Thant, M., & Nanjappa, K. A. (2006). Assessment of body condition in asian elephants (*Elephas maximus*). Zoo Biology, 25, 187–200.
- Wright, K. M., (2001). Restraint techniques and euthanasia. In K. M. Wright, & B. R. Whitaker (Eds.), *Amphibian medicine and captive husbandry* (pp. 111–122). Florida: Krieger Publishing Company.
- Wright, K. M., & Whitaker, B. R., (2001). Nutritional disorders. In K. M. Wright, & B. R. Whitaker (Eds.), Amphibian medicine and captive husbandry (pp. 73–87). Florida: Krieger Publishing Company.
- Yahnke, A. E., Grue, C. E., Hayes, M. P., & Troiano, A. T. (2012). Effects of the herbicide imazapyr on juvenile Oregon spotted frogs. *Environmental Toxicology and Chemistry*, 32, 228–235.

How to cite this article: Jayson S, Harding L, Michaels CJ, et al. Development of a body condition score for the mountain chicken frog (*Leptodactylus fallax*). *Zoo Biology*. 2018;37:196–205. https://doi.org/10.1002/zoo.21409